

# Converting rabbit hybridoma into recombinant antibodies with effective transient production in an optimized human expression system

Presented at HAH Belfast 2016 by

Dr. Tim Welsink

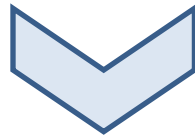
Molecular Biology  
Transient Gene Expression



# CASE STUDY

## RAB-MAB CONVERSION

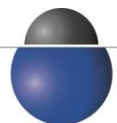
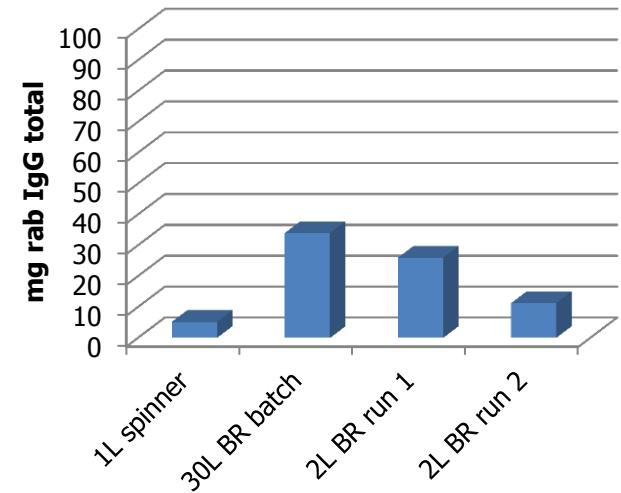
monoclonal rabbit hybridoma AB



recombinant rabbit AB produced by TGE

- 1) cDNA sequencing
- 2) Vector design, preparation
- 3) Transient production
- 4) Purification
- 5) QC

## Hybridoma rab. AB yield



# CASE STUDY

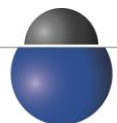
Sequence alignment HC variable region:

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AK6.P16-20.HC.K2 QSVEESGGRLVTPGTPLTLTCTVSGIDLSSYKMSWVRQAPGKGLEWIGVISRTGITIYAS 60
AK6.P16-20.HC.K3 QSVEESGGRLVTPGTPLTLTCTVSGIDLSSYKMSWVRQAPGKGLEWIGVISRTGITIYAS 60
AK6.P16-20.HC.K4 QSVEESRGRRLVTPGTPLTLTCTVSGIDLSSYKMSWVRQAPGKGLEWIGVISRTGITIYAS 60
AK6.P18-20.HC.K1 QSLI EESGGRLVTPGTPLTLTCTVSGIDLSSYKMSWVRQAPGKGLEWIGVISRTGITIYAS 60
AK6.P18-20.HC.K2 QSLI EESGGRLVTPGTPLTLTCTVSGIDLSSYKMSWVRQAPGKGLEWIGVISRTGITIYAS 60
AK6.P18-20.HC.K4 QSLI EESGGRLVTPGTPLTLTCTVSGIDLSSYKMSWVRQAPGKGLEWIGVISRTGITIYAS 60

AK6.P16-20.HC.K2 WAKGRFTISRTSTTVDLKITSPTTEDTATYLCARGDGGWGYGHIWGPGLVTIIS 116
AK6.P16-20.HC.K3 WAKGRFTISRTSTTVDLKITSPTTEDTATYLCARGDGGWGYGHIWGPGLVTIIS 116
AK6.P16-20.HC.K4 WAKGRFTISRTSTTVDLKITSPTTEDTATYLCARGDGGWGYGHIWGPGLVTIIS 116
AK6.P18-20.HC.K1 WAKGRFTISRTSTTVDLKITSPTTEDTATYLCARGDGGWGYGHIWGPGLVTIIS 116
AK6.P18-20.HC.K2 WAKGRFTISRTSTTVDLKITSPTTEDTATYLCARGDGGWGYGHIWGPGLVTIIS 116
AK6.P18-20.HC.K4 WAKGRFTISRTSTTVDLKITSPTTEDTATYLCARGDGGWGYGHIWGPGLVTIIS 116
```

Complementarity Determining Regions (CDRs) are colored, mismatches are highlighted

Primer sequences from  
Seeber et al., 2014 & Rader et al., 2000



# CASE STUDY

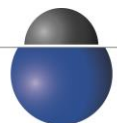
Sequence alignment LC variable region:

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AK6.P8-13.LC.K2  ELVLTQTPASVEAAVGGTVTIKCQASQSI SNLLAWYQOKPGQRPKLLIYYASNLAGVSS 60
AK6.P9-11.LC.K1  ELDLTQTPASVEAAVGGTVTIKCQASQSI SNLLAWYQOKPGQRPKLLIYYASNLAGVSS 60
AK6.P9-11.LC.K2  ELDMTQTPASVEAAVGGTVTIKCQASQSI SNLLAWYQOKPGQRPKLLIYYASNLAGVSS 60
AK6.P9-11.LC.K3  ELDLTQTPASVEAAVGGTVTIKCQASQSI SNLLAWYQOKPGQRPKLLIYYASNLAGVSS 60
AK6.P10-13.LC.K1 ELDLTQTPASVEAAVGGTVTIKCQASQSI SNLLAWYQOKPGQRPKLLIYYASNLAGVSS 60

AK6.P8-13.LC.K2  RFKGSGSGTEYTLTISGVQCADAATYYCQSYYYSSSTSNYAFDFGGGTEV VVK 112
AK6.P9-11.LC.K1  RFKGSGSGTEYTLTISGVQCADAATYYCQSYYYSSSTSNYAFDFGGGTELEI- 111
AK6.P9-11.LC.K2  RFKGSGSGTEYTLTISGVQCADAATYYCQSYYYSSSTSNYAFDFGGGTELEI- 111
AK6.P9-11.LC.K3  RFKGSGSGTEYTLTISGVQCADAATYYCQSYYYSSSTSNYAFDFGGGTELEI- 111
AK6.P10-13.LC.K1 RFKGSGSGTEYTLTISGVQCADAATYYCQSYYYSSSTSNYAFDFGGGTELEI- 111
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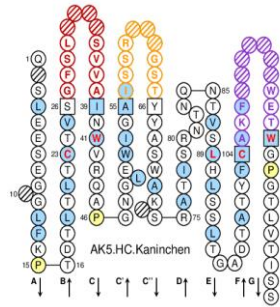
Complementarity Determining Regions (CDRs) are colored, mismatches are highlighted

Primer sequences from  
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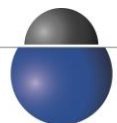
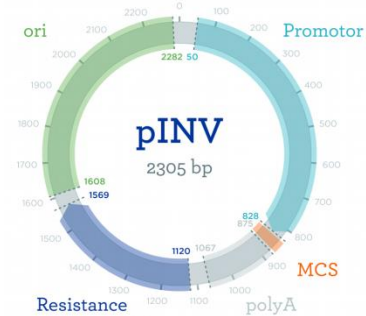
# CASE STUDY

- cDNA Design
- IgG gene synthesis
- Cloning into expression vector
- Low-endotoxin plasmid preparation
- Transient gene expression



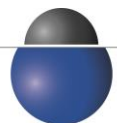
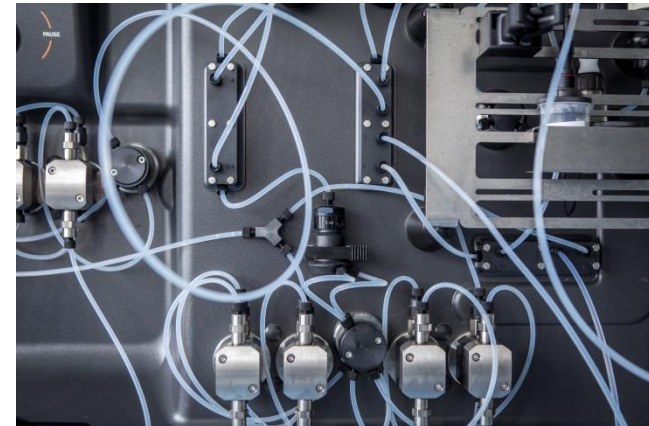
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GGC GGC CGC GAT ATC CCT AGG GGC ACC ATG AAG TGG GTC ACC TTT ATC TCC CTG CTG TTC CTG TTC
TCC TCC GCC TAC TCC GAA CTG GAC ATG ACC CAA ACC CCT GCC TCC GTG GAA GCT GCT GTT GGT GGT
ACT GTC ACC ATT AAG TGT CAA GCG AGT CAG AGC ATA TCC AAC CTG TTG GCC TGG TAT CAG CAG AAA
CCT GGG CAA AGR CCG AAA CTG CTC ATC TAT TAC GCC TCT AAT CTT GCT TCA GGC GTT TCC AGT GGG
TTC AAA GGG TCA GGA AGC GGT ACC GAG TAT ACA CTG ACC ATT TCT GGC GTC CAG TGT GCA GAT GCT
GCC ACC TAT TAC TGC CAG TCC TAC TAC TAC TCT AGC ACT TCC AAC TAT GCG TTC GAG TTT GGA GGA
GGG ACA GAA GTG GTG GTA AAG GGC GAT CCA GTG GCA CCC ACA GTC CTC ATC TTT CTT CCA GCA GCA
GAC CAG GTA GCC ACC GGT ACT GTC ACA ATC GTG TGT GTG GTC ACC AAC TAG TTT CCC GAC GTT ACC
GTG ACT TGG GAG GTT GAC GCC ACA ACT CAG ACC ACA GGC AIT GAG AAC AGC AAG ACA CCC CAG AAT
AGT GCC GAT TGC ACC TAT AAC CTG TCA AGC ACC CTT ACC CTG ACT TCA ACC CAG TAC AAT AGC CAC
AAG GAG TAC ACA TGC AAA GTG ACT CAA GGA ACC ACG TCT GTC GTG CAG AGC TTC AAT AGG GGC GAT
TGC TAA TAG GTT TAA ACT TAA TTA AAA GCT T
    
```



# CASE STUDY

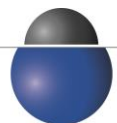
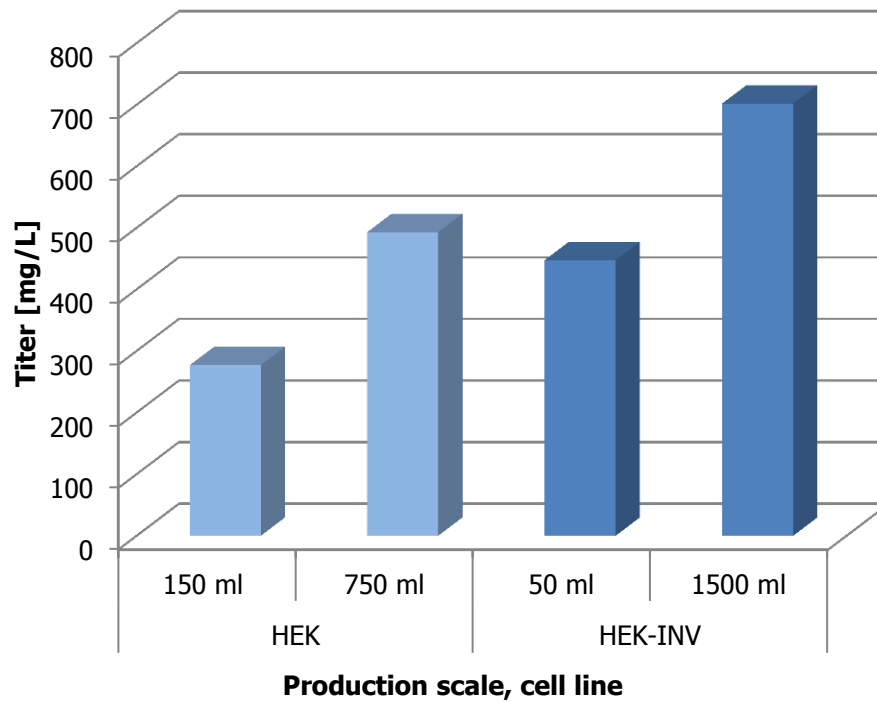
- Transient gene expression
  - Cell line: HEK or HEK-INV
  - Transfection protocol: optimized
  - Culture protocol: optimized
  - Scale: 50 to 1,500 ml
  - Production for 7 days
  - Harvest by centrifugation
  
- Purification
  - AF-rProtein A-650F (Tosoh Bioscience)
  - Äkta chromatography system
  - Elution by low pH
  - Dialysis to PBS pH 7.4



# CASE STUDY

## RESULTS

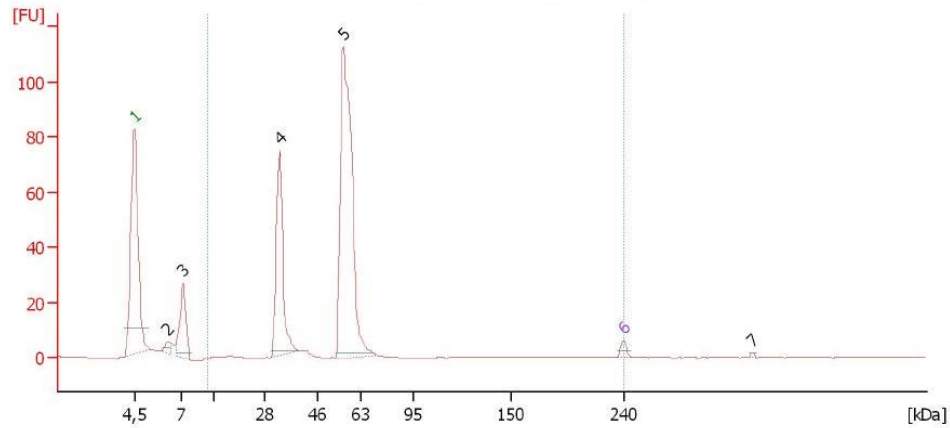
- 4 Batches produced
- Different production scale
- 2 cell lines



# CASE STUDY

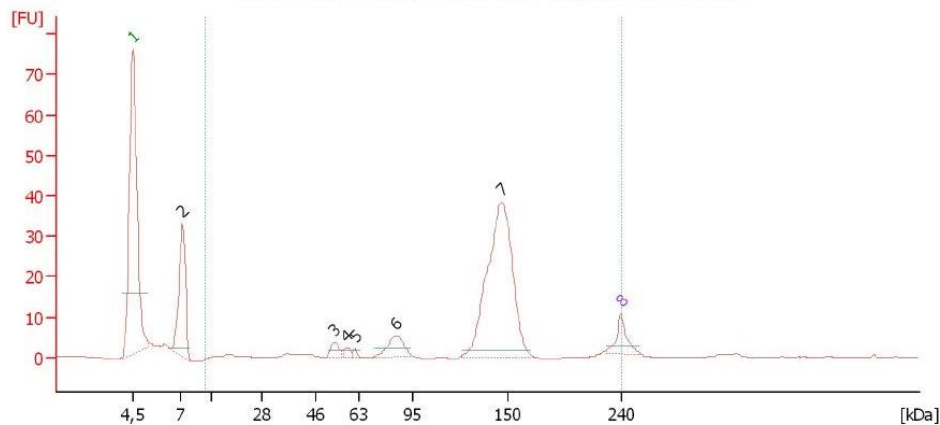
- QC: capillary gel electrophoresis for purity

**Reducing conditions**, Agilent 2100-BioAnalyzer, Protein 230 Kit

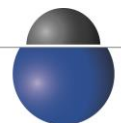


Purity  $\geq$  98%

**Non-reducing conditions**, Agilent 2100-BioAnalyzer, Protein 230 Kit



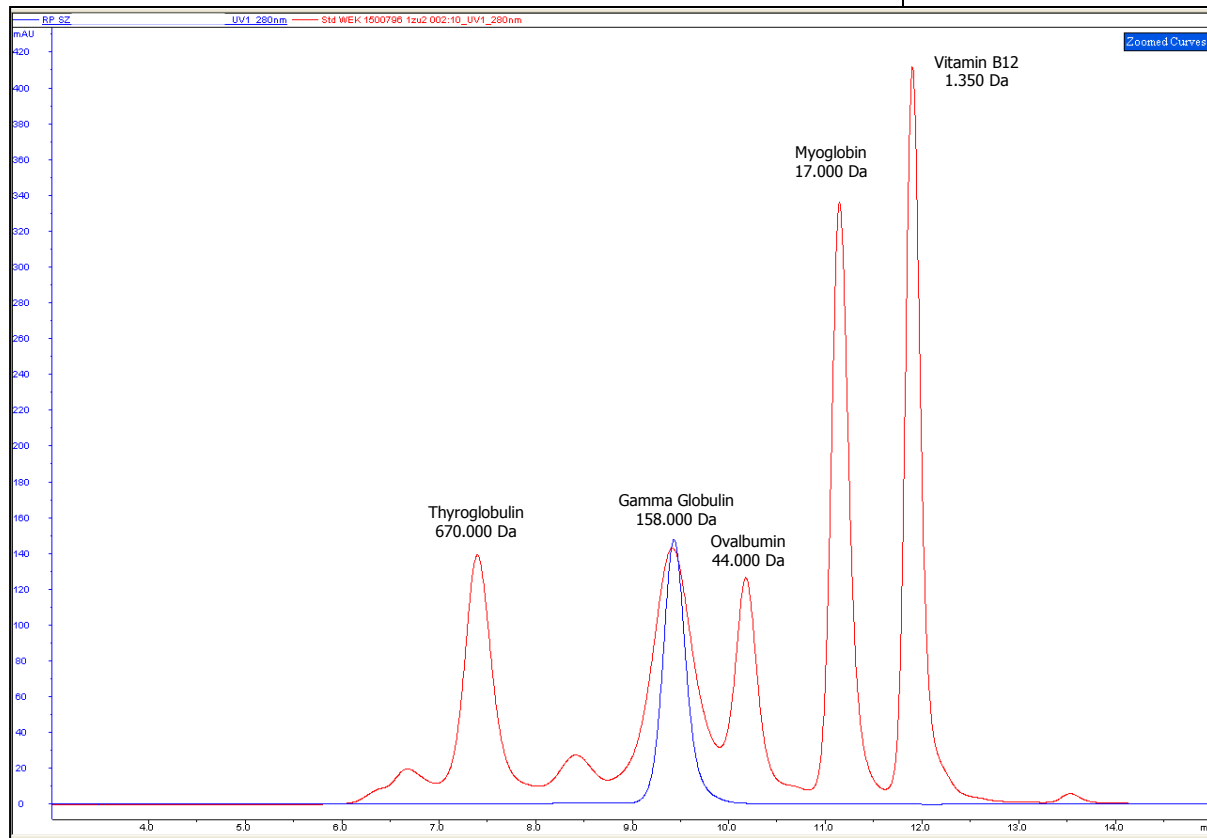
Purity  $\geq$  85%



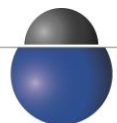


# CASE STUDY

- Analytical SEC for determination of aggregates

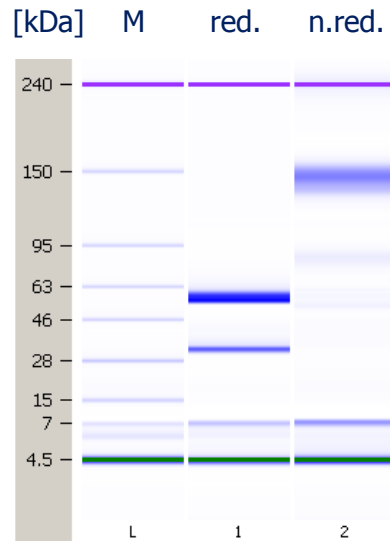


AppliChrom® ABOA-ProteSep S-L 5 $\mu$ , 300mm x 8mm

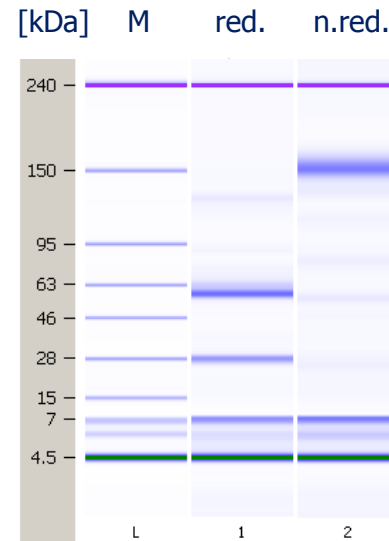


# CASE STUDY

## Recombinant rab-mab

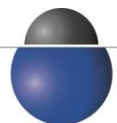
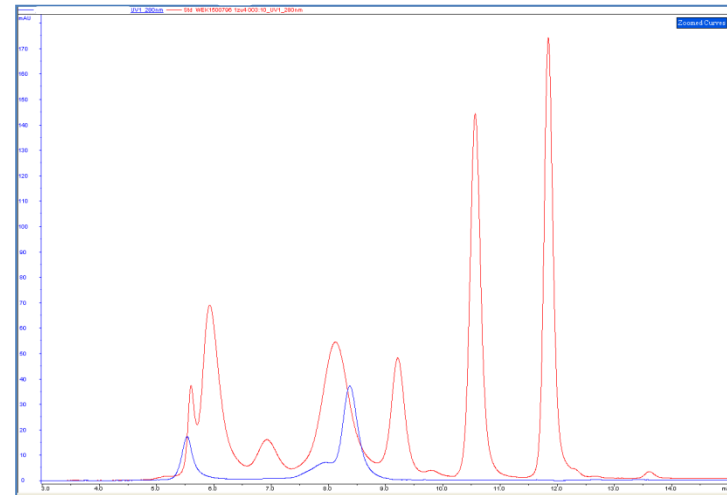
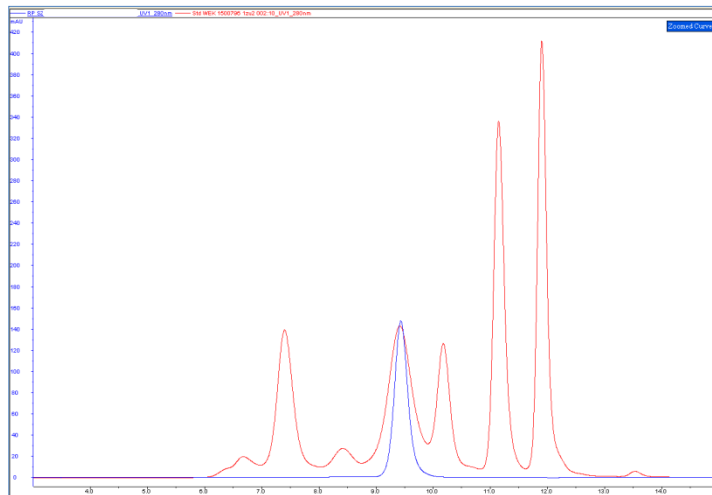


## Hybridoma rabbit AB



Right: CGE

Below: analyt. SEC



# Summary

- We had to address various needs to aim the goal
- Improvement of established methods
- New methods developed

Sequencing of Fv, production platform

- Case study rab-mab conversion & production
- Production and purification of 4 independent batches

Outstanding:

- Detailed results upon activity of the recombinant vs. hybridoma rab-mab

Conclusion:

New rab-mab production system allows to generate mg to g scales recombinant rabbit IgGs by transient gene expression within weeks.



# ACKNOWLEDGEMENTS



Dr. Wolfgang Weglöhner

Sebastian Püngel  
Penélope Soto Villegas  
Vanessa Vater

Dr. Sabrina Schindler  
Mohamad Eidi

Molecular Biology Lab  
Downstream Processing Lab

Cooperation partners:



# QUESTIONS



Please leave your question in our linkedin group [transient-transfection](#) or write use our [contact form](#)

